

Surface Enhanced Raman Scattering of Crystal Violet with Low Concentrations Using Self-Assembled Silver and Gold-Silver Core-Shell Nanoparticles

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ABSTRACT— The active substrates in surface enhanced Raman scattering (SERS) spectroscopy were prepared through self-assembly of nanoparticles on functionalized glasses. Colloidal silver nanoparticles (Ag NPs) were prepared chemically in two different sizes by reduction of AgNO_3 using trisodium citrate and sodium borohydride. Gold-silver core-shell nanoparticles were also prepared to compare between the optical behaviors of their silver shell and Ag NPs. Absorption spectra of nanoparticles were measured by ultraviolet-visible (UV-Vis.) spectroscopy, and their approximate sizes were determined by dynamic light scattering (DLS). The core-shell nanoparticles were approximately the same size as the largest Ag NPs (35 nm) and had the same maximum absorption wavelength. The potential of these substrates for detection applications was investigated with 1 M and 0.1 mM solutions of Raman-active molecule of crystal violet (CV) dye. The Raman enhancement signal was recorded for 0.1 mM solution with 532 nm laser wavelength, and the obtained spectra enhancement factor (EF) was calculated. EF values indicated that although the silver and gold-silver core-shell nanoparticles had the same surface plasmon resonance, the substrate with smaller Ag NPs had the highest enhancement factor compared to other substrates, which was 9.5×10^3 , and the core-shell substrate even had a slightly lower enhancement factor in compare with the large Ag NPs.

KEYWORDS: Crystal Violet, gold-silver core-shell nanoparticles, self-assembly, silver

nanoparticle, Surface-enhanced Raman Spectroscopy, SERS.

I. INTRODUCTION

Surface enhanced Raman scattering (SERS) spectroscopy is an analytical tool in the detection and characterization of chemical structures of molecules. There are several major factors that can contribute to the SERS enhancement effect, including: (1) electromagnetic enhancement resulting from collective free-electron oscillation, which enhances Raman signals of molecules adsorbed on the metal surface [1], and (2) charge-transfer between the metal nanoparticles and molecule (known as the chemical enhancement) [2-3].

Therefore, noble metal nanoparticles play a key role in achieving high sensitivity in SERS enhancement. In recent years, Au and Ag nanoparticles, due to their localized surface plasmon resonance (LSPR), have attracted attentions and play significant roles in SERS investigations. Au colloids are biocompatible and stable in comparison with Ag nanoparticles. These advantages make gold nanoparticles a viable option for using in SERS substrates. However, Ag nanoparticles show higher enhancement factors in visible spectrum [4-6]. Therefore, deposition of Ag shell on Au core can lead to further enhancement of Raman signal with significant properties of Au nanoparticles [7-8].

There are different techniques for reducing the metal shell to the surface of core particles [7,9]. The chemical reduction with citrate is used in this study. This method enables us to create a required number of nanoparticles easily and in a controlled manner.

There exist different methods of fabrication of good reproducible, low-cost, highly uniformed and efficient SERS substrates. Self-assembly of nanoparticles on surfaces of glass slides is one of the best choices to achieve uniformed SERS substrates with the ability to be reproduced [10-11].

In this study, self-assembly method was used to fabricate SERS substrates by Ag and Au@Ag NPs. The SERS activity of substrates assembled by Au@Ag and Ag NPs were compared for low concentrations of crystal violet molecule as an analyte.

II. EXPERIMENTAL PROCEDURES

A. Materials

Silver nitrate (AgNO_3 , 99%), hydrogen tetrachloroaurate (III) hydrate ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$, 99.9%), trisodium citrate dihydrate ($\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$, 99%), (3-aminopropyl) triethoxysilane (APTES, 99%), sulfuric acid (H_2SO_4), hydrogen peroxide (H_2O_2), ammonia water ($\text{NH}_3 \cdot \text{H}_2\text{O}$) and crystal violet were purchased from Merck Millipore. The stock solution was prepared by dissolving the crystals of the dye and preparing the aqueous solutions using de-ionized water.

B. Preparation of Silver Nanoparticles

Ag-1 nanoparticles were prepared using the following method. 30 ml of 2 mM NaBH_4 solutions were used in a cold bath as a reducing and stabilizing agent. 10 ml of 1 mM AgNO_3 were added to this solution while stirring over a period of 3 minutes. The solution eventually turned yellow indicating that the reaction was completed [12].

Ag-2 NPs were synthesized by the citrate reduction method and AgNO_3 solution (1 mM, 20 ml) as a metal salt precursor was heated. After boiling under vigorous stirring, 1.4 ml of

trisodium citrate (1 wt%) was injected into the solution. During the process, this solution was heated for another 10 minutes [13]. Then a green-gray colloid was removed from the heating device and stirred until cooled to reach room temperature.

C. Preparation of gold nanoparticles

Gold nanoparticles were synthesized according to the Frens method [14]. In order to do this, 20 ml of HAuCl_4 solution (0.01 wt%) was heated to boiling. Then, 0.4 ml of 1 wt% trisodium citrate aqueous solution was added to the boiling solution under vigorous stirring. When the color of mixture turned to wine-red, the heating was stopped.

D. Preparation of gold-silver core-shell nanoparticles

Au colloid was heated and then AgNO_3 solution (5 ml, 20 mM) and sodium citrate (5ml, 13.5 mM) simultaneously were added drop-by-drop into gold colloids under vigorous stirring. The mixture was continuously heated and stirred for 20 minutes, resulting in the formation of Au@Ag core-shell nanoparticles [15].

E. Preparation of Substrates

The glass slides were cleaned with DI water and ethanol successively under ultrasonication. Then, the glass slides were immersed in piranha solution ($\text{H}_2\text{SO}_4/\text{H}_2\text{O}_2=7:3$ v/v) and boiled at 80 °C for 30 minutes. After being washed with deionized water, they were immersed in solution of $\text{H}_2\text{O}_2/\text{NH}_3/\text{H}_2\text{O}$ (1:1:5, v/v/v) for 30 minutes at 75 °C. Next, the activated slides were immersed into an ethanol solution of APTES (1%) for 24 h to form a silane monolayer. Then, the glass slides were rinsed several times with ethanol to remove unbound silane and dried at 110 °C for 1 hour [16].

F. Self-Assembly Nanoparticles on Glass Slide

The amine-surface substrates were dipped into the prepared colloidal solutions. After 12 hours, the self-assembled nanoparticles slides were rinsed with deionized water and dried at

room temperature. By deposition of Ag-1, Ag-2, and Au@Ag NPs on the glass slides, three samples were prepared and named as sample-1, sample-2, and sample-3, respectively.

G. Sample Preparation for SERS

The aqueous solution of crystal violet at different concentrations were prepared. Desired low concentrations of crystal violet were dropped on the substrates. After evaporating the solvents, the SERS activities of the substrates were measured.

H. Characterization

UV-Visible absorption spectra of Ag, Au and Au@Ag colloidal solutions were recorded by a PG-Instruments model T80+ spectrometer with a 10 mm quartz cell. Baselines were recorded using milli-Q pure water and before measurements, the samples were diluted twice. The fluorescence emission spectrum was recorded using a spectrofluorometer (Gilden photonics) equipped with a xenon lamp. The size distribution of Ag, Au and Au@Ag NPs in the colloids were measured by a Zetasizer Nano-ZS system (Malvern Instruments). The measurements were performed at room temperature with a laser wavelength of 632.8 nm. Raman and SERS spectra were collected using a Nicolet Almega Dispersive Raman Spectrometer (Thermo Fisher Scientific Inc.) equipped with a maximum power of 100 mW, 532 nm laser. All spectra were collected at 10 mW of power, 32 accumulations.

III. RESULTS AND DISCUSSION

A. UV-Vis Absorption and Fluorescence Emission Spectra analysis

The optical properties of the nanoparticles were studied via UV-Visible spectroscopy. Fig. 1 shows the SPR maximum absorption, λ_{\max} , of silver, gold and gold-silver core-shell nanoparticles. The Ag-1 and Ag-2 colloids exhibit one SPR band around 392 and 428 nm, respectively.

The absorption peak at 521 nm belongs to the surface plasmon absorption of gold NPs. After the addition of silver salt solution, the plasmon

peak of the Au nanoparticles shifts toward the blue region ($\lambda_{\max} = 411$ nm). This blue shift reveals a change in the plasmonic properties of nanoparticles and indicates surface modification of Au NPs. The shape of the curve was not changed, which can be concluded that no aggregation of nanoparticles was occurred. The presence of only one peak in absorption band confirmed the formation of a uniform shell on the core particles in the colloid [17].

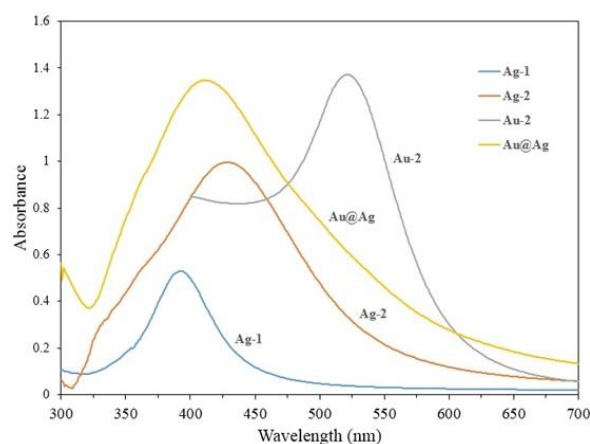


Fig. 1. UV-Vis absorption spectra of Ag-1, Ag-2, Au and Au@Ag nanoparticles.

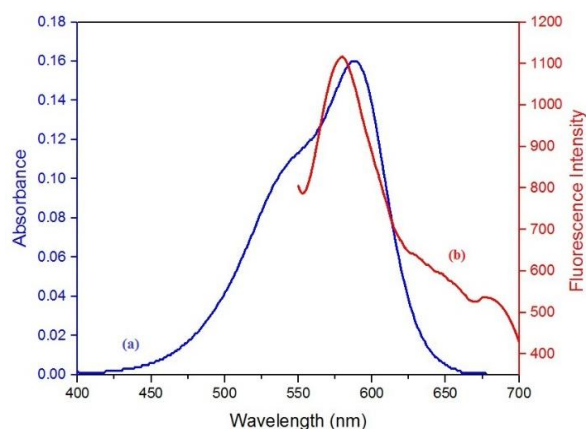


Fig. 2. (a) UV-Vis absorption and (b) fluorescence emission spectra of crystal violet.

Figure 2(a) shows the absorption spectra for crystal violet, that maximum absorbance occurred at 588 nm. Because the maximum absorbance of CV molecule is near excitation laser line (532 nm), the resonance has occurred. However, this overlap can increase the Raman intensities and decrease the detection limits, but a significant fluorescence background is revealed. This background can

substrates [20], the study showed that symmetry of CV solution is D_3 [21].

Figure 1 displays four histograms (a, b, c, d) showing the size distribution of AuNPs. The x-axis represents Size (d.nm) on a logarithmic scale from 1 to 1000. The y-axis represents Intensity (%) from 0 to 10. Histogram (a) shows a main peak at ~10 nm and a small peak at ~200 nm. Histogram (b) shows a main peak at ~30 nm and a small peak at ~10 nm. Histogram (c) shows a main peak at ~15 nm and a small peak at ~1 nm. Histogram (d) shows a main peak at ~30 nm.

Fig. 3. Measurement of particle size distribution of (a) Ag-1, (b) Ag-2, (c) Au, and (d) Au@Ag nanoparticles by DLS.

C. Raman Spectra Analysis

The diagram illustrates the chemical modification of a glass surface. The top part shows the APTES process: a glass surface with hydroxyl groups (OH) reacts with APTES to form a silane layer with terminal amino groups (NH₂). The middle part shows the 'Assembly of NPs' step, where nanoparticles (represented as grey spheres) are deposited onto the amino-functionalized surface. The bottom part is a magnified view of the final structure, showing the nanoparticles on the glass surface, with the APTES layer and its amino groups clearly visible.

Fig. 4. The diagram represents the preparation steps of glass functionalization via APTES and deposition of nanoparticles on the modified glass by immersing the slides in a colloidal nanoparticle.

The Raman spectra of the crystal violet molecule are shown in Fig. 6. Raman spectra of CV powder show that the CV peaks are very stronger than the solutions, whereas the intensity of signals in solutions are very weak and they can hardly be seen. The peak arising from the symmetric stretching vibration of C-

C appears at 1626 cm^{-1} in CV powder and 1 M solution and 1630 cm^{-1} in 0.1 mM solution, respectively. Another peak at 1549 cm^{-1} are assigned to the stretching vibration of $\text{C}_{\text{ring}}\text{N}$ and the symmetric bending of CH_3 . This peak does not appear in the SERS spectra. In addition, the peaks at 1391 and 1188 cm^{-1} in powder sample, are attributed to the C-H bending and the antisymmetric stretching of $\text{CC}_{\text{center}}\text{C}$, respectively. In the solutions, the peak intensities strongly decreased, so that in 0.1 mM concentration, most of the peaks were hardly characterized.

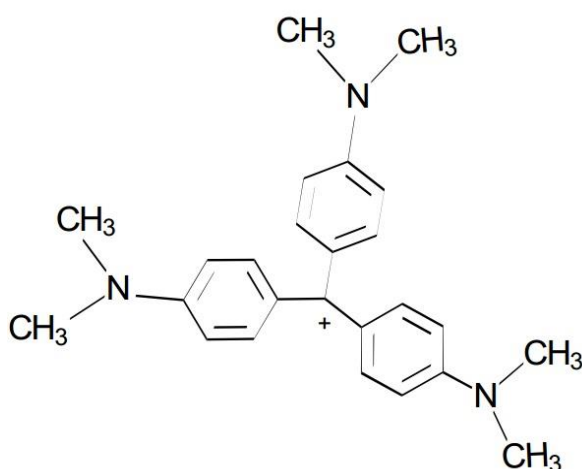


Fig. 5. Structure of crystal violet (CV).

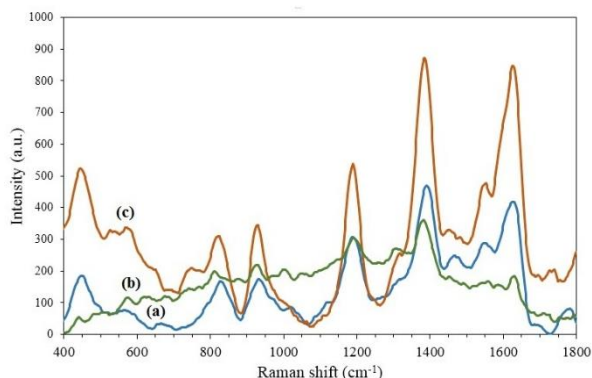


Fig. 6. Raman spectra of crystal violet (CV), (a): powder sample, and solutions with different concentrations including (b): 0.1 mM, and (c): 1 M.

After using the self-assembled substrates, SERS spectra are collected. Ag NPs with different particle sizes (samples-1, 2) and Au@Ag NPs (sample-3) were used as substrates to acquire SERS spectra of CV analyte with 0.1 mM concentration. The SERS spectra are shown in Fig. 7.

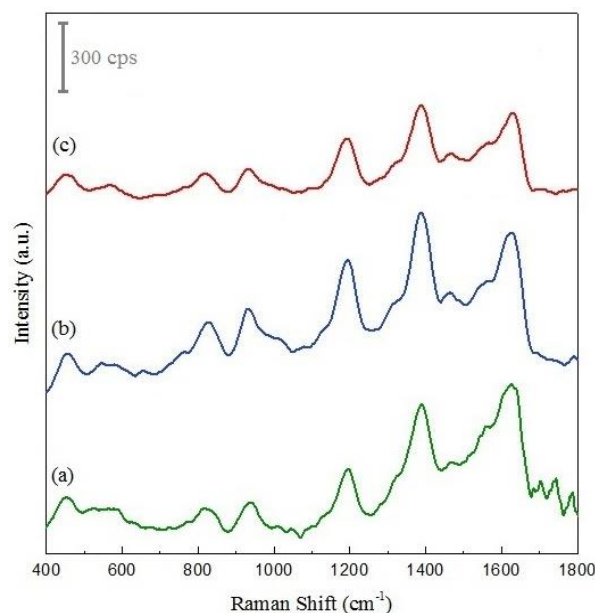


Fig. 7. SERS spectra of 0.1 mM aqueous solution of crystal violet (CV) adsorbed on (a): sample-1, (b): sample-2, and (c): sample-3.

The assignment peaks of SERS spectra are summarized in Table 1. These vibrational modes fall within the related wavenumbers reported in the literature [22]. The enhancement of Raman spectra of CV on the substrates can be demonstrated by calculating the related enhancement factors (EFs). The SERS EFs were calculated based on the most widely used equation:

$$EF = \frac{I_{SERS}/C_{SERS}}{I_{RS}/C_{RS}}$$

In the aforementioned equation I_{RS} corresponds to the Raman intensity obtained under non-SERS conditions at an analyte concentration of C_{RS} , and I_{SERS} corresponds to the Raman intensity obtained for the SERS substrate under a certain concentration of C_{SERS} . The experimental conditions, such as the laser wavelength, laser power, microscope objective or lenses, spectrometer, and measuring conditions on the substrate, are identical in all cases.

Based on the characteristic peak at 1626 cm^{-1} and using $C_{RS}=1\text{ M}$, $C_{SERS}=10^{-4}\text{ M}$ and the intensities obtained from Fig. 7, the EFs of substrates were estimated to be about 9.5×10^3 ,

9.2×10^3 , and 4.8×10^3 corresponding to the sample-1, sample-2, and sample-3, respectively.

Table 1. Peak assignments of crystal violet (CV) measured in SERS spectra, (v: stretching vibration (s: symmetric; as: antisymmetric; δ : bending vibration; ρ : rocking; γ : out-of-plane deformation)

Assignment	$\nu_{SERS} (cm^{-1})$		
	Sample-1	Sample-2	Sample-3
$\nu_s(C-C)$	1626	1628	1630
$\delta_{as}(CH_3)$	1468	1464	1468
$\delta(CH)/\delta_s(CH_3)/\delta(CCC)_{ring}$	1389	1387	1387
$\nu_{as}(CC_{center}C)$	1196	1194	1192
$\rho(CH_3)/\nu(CN)$	935	932	934
$\delta(CC_{center}C)$	814	827	820
$\gamma(CCC)/\delta(CNC)/\delta(CC_{center}C)$	563	579	567
$\delta(CNC)$	453	453	449

Enhancement factors indicated that all three substrates were well able to reveal very low concentrations. Because of the similarity of the absorption spectra of larger Ag NPs and the gold-silver core-shell NPs, it was expected that sample-2 and sample-3 behave in the same way as Raman signal enhancement, but the calculated EF values differed significantly, and Ag NPs performed better in Raman signal enhancement. This is due to the electromagnetic fields decay exponentially in metals, and that core-shell NPs only consist of a thin outer shell of silver on gold NPs, and thus do not have the same optical properties of pure Ag NPs with the same size.

The optical absorption of silver is the reason for the high signal enhancement by the silver substrates, because interband transitions play a key role in the intensity of SERS. The interband transition of silver metal occurs in the UV region, therefore its absorption at the visible Raman laser wavelength, 532 nm, results in high Raman signal intensities [23].

The results of sample-1 and sample-2 are not in full agreement with theoretical predictions and experimental data, because it was expected that as the size of Ag NPs increases,

the electromagnetic field enhances and the stronger surface plasmons provides higher enhancement. But it should be also considered with self-assembly on the surface of the glass, nanoparticles tend to aggregate and as a result, their size will increase. Thus, the small nanoparticles approach to the optimal size to achieve high Raman signal intensities [24]; In contrast, the ability of larger nanoparticles (sample-2) in detecting will decrease.

IV. CONCLUSION

Self-assembly of nanoparticles on glass is an efficient, low-cost, and easy method to prepare SERS substrates. To this end, substrates of gold-silver core-shell nanoparticles with an approximate diameter of 39 nm with Ag NPs in two different sizes (8 and 35 nm) were synthesized. Their function in detecting Raman modes using CV pigment was investigated and compared. The results showed that in the same experimental conditions, such as frequency and laser power, the strongest signal was recorded by the smallest Ag NPs substrate. All three substrates well detected Raman modes of 0.1 mM of CV solution, and the highest EF (9.5×10^3) is related to the Ag NPs with the smallest size.

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